

2019-nCoV and Influenza A+B Antigen Device (2–30°C)

CATALOGUE NUMBER	KIT SIZE (TESTS)
RADCOV5	20 Tests

Intended Use:

The 2019-nCoV and Influenza A+B Antigen Device is a rapid chromatographic immunoassay for the qualitative detection of antigens of SARS-CoV-2 and Influenza A and B viruses in nasal swab samples to aid in the differential diagnosis of COVID-19 and Influenza both of which characteristically display respiratory symptoms. For in vitro diagnostic use by trained professionals only.

Summary:

Coronaviruses are a large family of viruses that cause disease ranging from common cold symptoms to more severe pneumonia. They are enveloped, single strand RNA viruses. Coronaviruses are zoonotic, they can be transmitted from animals to humans. Existing examples include the Middle East Respiratory Virus (MER-CoV) and Severe Acute Respiratory Syndrome (SARS-CoV). Symptoms include high temperature, cough and breathing difficulties. In immunocompromised individuals symptoms can be more severe leading to pneumonia, severe acute respiratory syndrome or death.

Three types of influenza virus infect humans, A and B being the most common. These viruses are highly infectious being spread as the live virus in airborne water droplets from coughing and sneezing. Symptoms include fever, rhinorrhoea, sore throat, pain in muscles and joints, headache and fatigue. Effects can be severe in the lungs resulting in viral pneumonia and death, can lead to secondary disease such as bacterial pneumonia and exacerbate previous health conditions including asthma and heart failure.

Test Principle:

The test device operates as a double antibody immunoassay in two separate channels. In the first channel anti-SARS-CoV-2 antibody is immobilized on the membrane in the test zone. Particles conjugated with anti-SARS-CoV-2 antibody are coated on the membrane near the sample well. During the test nasopharyngeal swab extract is added to the sample well where it interacts with the antibody coated particles and SARS-CoV-2 antigens present in the sample will bind to the antibody. The antigen-particle complexes migrate up the membrane by capillary action where they interact with the anti-SARS-CoV-2 antibody at the Test line and are captured. A positive result is indicated when a coloured line forms at the Test line. The absence of any line development at the test zone indicates a negative result. To serve as a procedural control, a coloured line should always appear at the control line area indicating that proper volume of specimen has been added and membrane wicking has occurred.

In the second channel anti-influenza A and anti-influenza B antibodies are immobilized on the membrane at the A and B test lines respectively. Particles conjugated with anti-influenza A or anti-influenza B are coated on the membrane near the sample well. The extracted swab sample is added to the sample well where it interacts with the particles and influenza antigens present in the sample will bind to either or both antibodies. The antigen-particle complexes migrate up the membrane by capillary action where they interact with the anti-influenza A and/or anti-influenza B antibodies at the respective test lines and are captured. A positive result is indicated when a coloured line forms at the A and/or B test lines. The absence of any line development in the test zone indicates a negative result. To serve as a procedural control, a coloured line should always appear at the control line area indicating that proper volume of specimen has been added and membrane wicking has occurred.

Reagents:

The test device contains anti-SARS-CoV-2, anti-influenza A and anti-influenza B antibodies.

Materials Provided

Individually pouched test devices
Extraction Buffer
Extraction tubes
Swabs
Instructions for Use sheet

Materials not provided:

Timer

Precautions:

Follow Good Laboratory Practice procedures where samples and kits are handled and treat the device and all samples as if potentially infectious. Follow local regulations for correct disposal of samples.

Wear protective clothing including laboratory coat, disposable gloves and safety glasses when conducting the test.

IMPORTANT: Viral Transport Media (VTM) can interfere in the test results. Do not store swab samples in VTM.

Humidity and temperature can adversely affect results.

Storage and Stability:

The kit can be stored at room temperature or refrigerated (2 - 30°C). The test device is stable up to the expiry date printed on the sealed pouch. The device must remain in the sealed pouch until use. Do not freeze. Do not use after the expiry date.

Sample Collection and Storage:

The 2019-nCoV and Influenza A+B Antigen Device test is performed using an extracted nasopharyngeal swab sample.

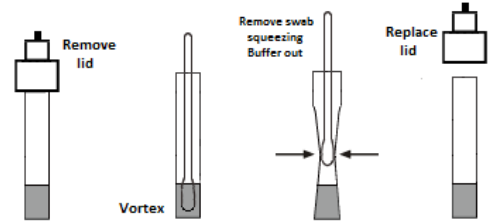
Collection: Insert the swab into the nasal cavity of the subject and push to the back of the nasopharynx. Wipe the swab over the posterior nasopharynx then withdraw the swab slowly whilst rotating it.

Transport and storage: Samples should be used in the test as soon as possible. If not tested

Immediately, the swabs should be tightly sealed in a dry specimen container, under which conditions they may be stored up to 8 hours at room temperature (15 – 30°C) or 24 hours at 2 - 8°C.

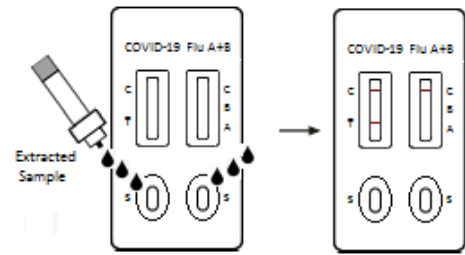
Sample Preparation:

- Use the Extraction Tubes and Extraction Buffer supplied in the kit.
- Line up Extraction Tubes in the test tube rack and add 350 µl Extraction Buffer. Insert the dry swab sample into the buffer and, holding the swab head below the surface of the buffer, stir the swab vigorously in the liquid to release antigens into the Extraction Buffer.
- Press the swab against the wall of the tube then lift out the swab while squeezing the sides of the tube around it to release as much Buffer as possible from the swab head.
- Fit the lid of the Extraction Tube securely. See illustration below.
- The extracted sample is stable for 2 hours at room temperature (15 – 30°C) or 24 hours at 2 - 8°C.



Procedure:

- Bring extracted samples, controls and the sealed test device to room temperature (15 – 30°C).
- Remove the test device from the sealed pouch, place it on a clean and level surface and use it immediately (but no later than one hour after opening).
- Tip up the Extraction Tube and dispense 3 drops of extracted sample (approximately 100 µl) into each of the Sample Wells. Start the timer.
- Wait for coloured lines to appear. Read the results at 15 minutes. Do not interpret any result after 20 minutes.



Interpretation of Results:

SARS-CoV-2 Positive: Two clear coloured lines appear in the left-hand reading window. One band appears at the control line (C) and one band develops at the Test line (T). This result indicates detection of SARS-CoV-2 antigens.

Influenza A Positive: Two clear coloured lines appear in the right-hand reading window. One band appears at the control line (C) and one band develops at the Influenza A test line (A). This result indicates detection of Influenza A antigen.

Influenza B Positive: Two clear coloured lines appear in the right-hand reading window. One band appears at the control line (C), and one band develops at the Influenza B test line (B). This result indicates detection of Influenza B antigen.

Influenza A and Influenza B Positive: Three clear coloured lines appear in the right-hand reading window. One band appears at the control line (C), one band develops at the Influenza A test line and one band develops at the Influenza B test line. This result indicates detection of both Influenza A and Influenza B antigens.

* NOTE: The intensity of colour development at the test lines will vary depending on the concentration of antigens present in the sample. Therefore, any shade of colour developing at the test lines should be considered positive.

Negative: One coloured line appears at the control line (C). No visible coloured lines appear in the test zones.

Invalid:



A Control line fails to appear. Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test with a new test device. If the problem persists, discontinue using the test kit immediately and contact your local distributor.

Quality Controls:

A procedural control is included in the test. A coloured line appearing in the control region (C) is the internal procedural control. It confirms sufficient specimen volume and correct procedural technique. Quality Controls are not supplied with this kit.

Limitations of the Test:

The Assay Procedure and the Assay Result Interpretation must be followed closely when testing for the presence of antigens to SARS-CoV-2 and Influenza A and B from individual subjects. Failure to follow the procedure may give inaccurate results. Swab samples stored in viral transport medium cannot be used in the 2019-nCoV and Influenza A+B Antigen Device. Samples extracted by a procedure in readiness for PCR testing cannot be used in this device.

The 2019-nCoV and Influenza A+B Device is limited to the qualitative detection of SARS-CoV-2, Influenza A and Influenza B antigens in extracted swab samples to aid in the differential diagnosis of COVID-19 and Influenza. The intensity of the test bands do not have a linear correlation with antigen concentration in the samples. A negative result for an individual subject indicates absence of a detectable level of SARS-CoV-2, Influenza A and Influenza B antigens but a negative test result does not preclude the possibility of exposure to or infection with these viruses. If symptoms persist a repeat of the test with a new swab sample is recommended after 2 – 4 days or test by an alternative method such as RT-PCR.

If blood is present in the swab sample or excess mucus this can lead to false positive results and false negative results can be obtained following incorrect sample collection, extraction or storage. The results obtained with this test should not be used as the sole criterion for diagnosis of SARS-CoV-2 or Influenza infection but be used in conjunction with other diagnostic procedures and clinical findings.

Performance Characteristics:

The 2019-nCoV and Influenza A+B Antigen Device has been evaluated in clinical trials testing samples from individuals displaying and not displaying symptoms. The reference method for the study was RT-PCR, the matched sample results for which were regarded as the gold standard, either positive or negative.

Results for SARS-CoV-2 antigen

2019-nCoV and Influenza A+B Antigen Device	Method	PCR		Total Results
	Results	Positive	Negative	
	Positive	80	1	
	Negative	3	120	
Total Results		83	121	204

Sensitivity: 96.4% (95% CI*: 89.8% - 99.2%) *Confidence Intervals

Specificity: 99.2% (95% CI*: 95.5% - 99.9%)

Accuracy: 98.0% (95%CI*: 95.1% - 99.5%)

Results for Influenza A and Influenza B

2019-nCoV and Influenza A+B Antigen Device	Method	Influenza A		Influenza B	
		PCR		PCR	
	Results	Positive	Negative	Positive	Negative
	Positive	38	2	39	2
	Negative	2	215	3	213
Total Results		40	217	42	215

Influenza A

Sensitivity: 95.0% (95%CI*: 82.6% - 99.5%) *Confidence Intervals

Specificity: 99.1% (95% CI*: 96.5% - 99.9%)

Accuracy: 98.4% (95% CI*: 95.9% - 99.5%)

Influenza B

Sensitivity: 92.9% (95%CI*: 80.3% - 98.2%)

Specificity: 99.1% (95% CI*: 96.5% - 99.9%)

Accuracy: 98.1% (95%CI*: 95.4% - 99.3%)

Precision

Seven levels of SARS-CoV-2 and Influenza A and B standard material including a Negative, a Weak SARS-CoV-2 antigen, a Strong SARS-CoV-2 antigen, a Weak Influenza A antigen, a Strong Influenza A antigen, a Weak Influenza B antigen, and a Strong Influenza B antigen were used to determine Intra and Inter assay precision. Ten replicates of each level were tested each day on 3 consecutive days. Results for each replicate for each level were correct, according to the designated result for the standard sample, with greater than 99% accuracy.

Cross-reactivity

The 2019-nCoV and Influenza A+B Antigen Device has been assessed for cross reactivity by testing specificity with a range of viruses associated with fever, cough and other respiratory symptoms and via cross reactivity caused by other pathogenic organisms.

Specificity with a range of viruses

Results for SARS-CoV-2

Virus	Dilution of virus in sample tested
Adenovirus type 3	3.16 x 10 ⁴ TCID50/ml
Adenovirus type 7	1.58 x 10 ⁵ TCID50/ml
Human Coronavirus OC43	2.45 x 10 ⁶ LD50/ml
Influenza A H1N1	3.16 x 10 ⁵ TCID50/ml
Influenza A H3N2	1 x 10 ⁵ TCID50/ml
Influenza B	3.16 x 10 ⁶ TCID50/ml
Human Rhinovirus 2	2.81 x 10 ⁴ TCID50/ml
Human Rhinovirus 14	1.58 x 10 ⁶ TCID50/ml
Human Rhinovirus 16	8.89 x 10 ⁶ TCID50/ml
Measles	1.58 x 10 ⁴ TCID50/ml
Mumps	1.58 x 10 ⁴ TCID50/ml
Parainfluenza virus 2	1.58 x 10 ⁷ TCID50/ml
Parainfluenza virus 3	1.58 x 10 ⁸ TCID50/ml
Respiratory syncytial virus	8.89 x 10 ⁴ TCID50/ml

Results for Influenza A and B

Virus	Dilution of virus in sample tested
Adenovirus type 3	3.16 x 10 ⁴ TCID50/ml
Adenovirus type 7	1.58 x 10 ⁵ TCID50/ml
Human Coronavirus OC43	2.45 x 10 ⁶ LD50/ml
Human Rhinovirus 2	2.81 x 10 ⁴ TCID50/ml
Human Rhinovirus 14	1.58 x 10 ⁶ TCID50/ml
Human Rhinovirus 16	8.89 x 10 ⁶ TCID50/ml
Measles	1.58 x 10 ⁴ TCID50/ml
Mumps	1.58 x 10 ⁴ TCID50/ml
Parainfluenza virus 2	1.58 x 10 ⁷ TCID50/ml
Parainfluenza virus 3	1.58 x 10 ⁸ TCID50/ml
Respiratory syncytial virus	8.89 x 10 ⁴ TCID50/ml

None of the virus samples caused any trace of colour line development at the test line regions indicating no cross reactivity of these viruses in the 2019-nCoV and Influenza A+B Antigen Device.

Cross reactivity with other pathogenic organisms

A range of bacterial and fungal pathogens were tested as samples at 1 x 10⁸ organisms/ml in the 2019-nCoV and Influenza A+B Antigen Device.

Arcanobacterium	Pseudomonas aeruginosa
Candida albicans	Staphylococcus aureus subsp. aureus
Corynebacterium	Streptococcus pneumoniae
Escherichia coli	Streptococcus epidermidis
Moraxella catarrhalis	Streptococcus pyogenes
Neisseria lactamica	Streptococcus salivarius
Neisseria subflava	Streptococcus sp group F

None of the organisms caused any trace of colour line development at the test line regions indicating no cross reactivity of these organisms in the 2019-nCoV and Influenza A+B Antigen Device.

References:

- Williams KM et al. Rapid diagnostic testing for URIs in children. Impact on physician decision making and cost. Infect. Med 2002; 19:109-111.
- Betts RF; 1995. Influenza virus. 1546-1567. In Principle and practice of infectious diseases. Mandell GF, Douglas RG and Bennett JE (eds) 4th ed Churchill Livingstone, Inc, New York.
- World Health Organisation Recommendations on the use of rapid testing for influenza diagnosis, WHO 9 July 2005.

Glossary of Symbols:

	Catalogue number		Temperature limitation
	Consult instructions for use		Batch code
	In vitro diagnostic medical device		Use by date
	Manufacturer		Do not reuse

